TWO NEW TERTIARY INDOLE ALKALOIDS OF STRYCHNOS DECUSSATA

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ABSTRACT.-From the chloroform fraction of the stem bark of Strychnos decussata (Pappe) Gilg (Loganiaceae), two new tertiary indole alkaloids, 10-hydroxyakagerine (1) and akagerinelactone (3), have been isolated. The structures of the alkaloids have been elucidated by means of their spectral data, particularly ¹H-nmr and ¹³C-nmr.

In a recent publication, we reported the isolation of five tertiary indole alkaloids, three of which were new, from the stem bark of *Strychnos decussata* (1). In continuation with our studies of the convulsant and muscle-relaxant constituents of S. decussata, we have now isolated two more indole alkaloids from the chloroform fraction of the stem bark.

RESULTS AND DISCUSSION

The proposed structures are presented in the figure.

Alkaloid 1 analyzed for $C_{20}H_{24}N_2O_3$ (M⁺, 340.1795); $C_{20}H_{24}N_2O_3$ requires 340.1787. The alkaloid's uv spectrum, which was strikingly similar to that of 10hydroxy-17-O-methylakagerine, showed a bathochromic shift on addition of alkali, suggesting the presence of a phenolic function. The ir spectrum showed absorption peaks at 3280 cm⁻¹ (-OH) and at 1665 cm⁻¹ (-C=O). A comparison of the ms of alkaloid 1 with that of akagerine (2) showed a constant difference of 16 mass units higher values for the fragments containing the aromatic moiety in alkaloid 1. The ¹H-nmr spectrum of alkaloid 1 exhibited signals for an aldehydic proton at $\delta 9.22$ (1H, s), an ethylidene chain at $\delta 6.52$ (1H, q) and $\delta 2.04$ (3H, d), and three aromatic protons in the region $\delta7.10-6.62$. This is almost identical with the ¹H-nmr spectrum of 10-hydroxy-17-O-methylakagerine, except for the absence of



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a three-proton singlet for a methoxy group in $\mathbf{1}$, suggesting that alkaloid $\mathbf{1}$ is 10hydroxyakagerine.

Alkaloid 3 analyzed for $C_{20}H_{22}N_2O_3$ (M⁺, 338.1655); $C_{20}H_{22}N_2O_3$ requires 338.1657. The uv spectrum of this alkaloid was typical of akagerine-like compounds (1). The ir spectrum of alkaloid **3** showed a strong absorption at 1735 cm⁻¹, suggesting the presence of an $\alpha_{\beta}\beta$ -unsaturated lactone. The ¹H-nmr spectrum of alkaloid 3 was very similar to that of akagerine, differing only in the protons of the side chain, H-18, H-19 and H-21. The signals due to the aldehydic proton, $\delta 9.24$ (1H, s), and the ethylidene chain protons, $\delta 6.43$ (1H, g) and $\delta 2.03$ (3H, d), present in the ¹H-nmr of akagerine were replaced by a two-proton triplet and a one-proton multiplet resonating at $\delta 4.80$ and $\delta 7.12$, respectively, in the spectrum of alkaloid 3. The assignments of the spectrum were carried out by decoupling experiments, further supporting a lactone ring in alkaloid 3. Irradiating the signals at $\delta 4.80$ and at $\delta 7.12$ revealed coupling between H-18 and H-19. Further decoupling studies showed long range coupling both between $\delta 3.45$ (H-15) and $\delta7.12$ (H-19) and between $\delta3.45$ and $\delta4.80$ (H-18). The ¹³C-nmr spectrum of alkaloid 3 was also similar to the spectrum of akagerine-like compounds, except for the carbons in the side chain. The signals for C_{18} (CH₃, 15.1), C_{19} (CH, 150.7), C_{20} (C, 147.8), and C_{21} (H-C=O-, 195.0) in akagerine disappeared in the ¹³C-nmr spectrum of alkaloid 3; the following signals were observed instead: CH_2 (70.23), CH (144.11), C (138.21) and C (173.40), respectively. The signals of the ¹H-nmr and ¹³C-nmr of alkaloid 3 are in full accord with the replacement of the ethylidene-aldehyde side chain in akagerine by a lactone ring in alkaloid 3, for which the name akagerinelactone has been given.

Further evidence for the presence of a lactone ring was provided by positive color reactions with reagents for the detection of an unsaturated lactone group.

Comparing the values of the ¹H-nmr and ¹³C-nmr spectra of akagerinelactone and akagerine, it is most likely that the configuration at the asymmetric carbons C-3, C-15 and C-17 are the same in the two alkaloids.

No pharmacological testing of the two new alkaloids was performed due to the limited amount of material isolated.

EXPERIMENTAL³

Details of plant collection, extraction and chromatographic procedures have been reported elsewhere (1).

10-HYDROXYAKAGERINE (1).—Purified by preparative tlc using cyclohexane-chloroform-diethylamine (50:40:10) as mobile phase, 10-hydroxyakagerine was obtained as a pale brown powder (8.5 mg) mp 154-156°. It gave uv: λ max (log ϵ) (ethanol): 230.5 (4.38), 280 (3.93), 308 (sh, 3.60) and 312 (3.52) nm; λ max (log ϵ) (addition of KOH): 232 (4.74), 279 (3.89) and 322 (3.54) nm; ir: ν max (KBr): 3280, 2900, 2820, 1665, 1620, 1460, 1360, 1210, 1150, 1100, 900, 830 and 785 cm⁻¹; ms: m/z (\mathcal{C}_0): 340 (11, M⁺), 322 (17), 293 (7), 276 (5), 253 (14), 214 (7), 202 (16), 201 (100), 200 (27), 187 (10), 172 (11), 160 (6), 159 (6), 144 (5) and 143 (2); ¹H-nmr (CDCl_s): 89.22 (1H, s, H-21), 7.10 (1H, d, J=9Hz, H-12), 6.70 (1H, d, J=2Hz, H-9), 6.62 (1H, dd, J= 9Hz, 2Hz, H-11), 6.52 (1H, q, J=7Hz, H-19), 6.04 (1H, broad s, H-17), 2.48 (3H, s, N-CH_s) and 2.04 (3H, d, J=7Hz, H-18) ppm.

ferric (III) chloride [4].

^{&#}x27;The uv spectra were run on a Shimadzu MPS-5000 UV-VIS spectrophotometer, the ir ³The uv spectra were run on a Shimadzu MPS-5000 UV-VIS spectrophotometer, the ir spectra on a Jasco-IRA-I-spectrophotometer and the ms spectra on an LKB 9000 instrument at 70 eV with direct inlet system. ¹H-mmr spectra were run on a Jeol 100 MHz spectrometer and the ¹³C-mmr spectrum was recorded on a Varian 100 MHz instrument at 25.2 MHz in the Fourier transform mode. Melting points were determined with a Leitz mikroskopheitztisch 350. Preparative thin-layer chromatography (tlc) was carried out on either precoated silica gel plates (0.25 mm, silica gel Si60 F₂₅₄, E. Merck) or on a 0.50 mm layer of silica gel (type 60, GF₂₅₄, E. Merck) spread on 20 x 20 cm glass plates. Reagents used for the detection of unsaturated lactones on tlc: Keddes reagent [2], Bal-jets reagent [2], Raymonds reagent [3], Legals reagent [4] and hydroxyl ammonium chloride: ferric (III) chloride [4].

AKAGERINELACTONE (3).—On purification by preparative tlc using diethyl ether-ethanol-diethylamine (90:3:7) as mobile phase, *akagerinelactone* crystallized from ethanol/diethyl ether as colorless needles (17 mg), mp 184–186°. It gave uv: λ max (log ϵ) (ethanol) 222 (4.33), 276 (3.90), 283 (3.91) and 293 (3.80) nm; ir: ν max (KBr): 3350, 3000, 2900, 2820, 2760, 1735, 1640, 1660, 1460, 1380, 1350, 1310, 1270, 1200, 1190, 1090, 1060, 1040, 980, and 735 cm⁻¹: ms: m/z (C_c): 338 (9, M⁻¹), 323 (4), 320 (7), 214 (6), 198 (8), 186 (15), 185 (100), 183 (10), 171 (18), 169 (5), 156 (13), 144 (8) and 143 (6): ¹H-nmr: (CDCl₂+CD₃OD): $\delta 7.58-7.16$ (4H, m, H–9, H–10, H–11, H–12), 7.12 (1H, m, H–19), 6.30 (1H, m, H–17), 4.80 (2H, t, J=1Hz, H–18) and 8.45 (1H, m, H–15) and 2.59 (3H, s, N–CHz) ppm: ¹³C–nmr: (CDCl₃+CD₃OD) C–2 136.11,* C–3 60.04, C–5 50.58, C–6 19.63, C–7 108.38, C–8 126.34, C–9 118.39, C–10 121.79, C–11 119.68, C–12 108.68, C–13 135.40*, C–14 38.81, C–15 29.06, C–16 36.66, C–17 75.04, C–18 70.23, C–19 144.11, C–20 138.21, C–21 173.40 and N–CHz 42.15 ppm. ARAGERINELACTONE (3).—On purification by preparative the using diethyl ether-ethanol-

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LITERATURE CITED

- W. N. A. Rolfsen, A. A. Olaniyi and P. J. Hylands, J. Nat. Prod., 43, 97 (1980).
 H. H. S. Fong, M. Tin-Wa and N. R. Farnsworth, *Phylochemical Screening*, Dept. of Phar-
- macognosy and Pharmacology, Univ. of Illinois at the Medical Center, Chicago, Illinois. E. Stahl, Dünnschichtchromatographie, Springer Verlag, Berlin, Heidelberg, New York 3. (1967).
- 4. Anfarbereagenzien für Dünnschicht- und Papier Chromatographie, E. Merck, Darmstadt.

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